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ARTICLE OPEN

Regulation of amino-acid metabolism controls flux to lipid accumulation in *Yarrowia lipolytica*Eduard J Kerkhoven¹, Kyle R Pomraning², Scott E Baker² and Jens Nielsen^{1,3}

Yarrowia lipolytica is a promising microbial cell factory for the production of lipids to be used as fuels and chemicals, but there are few studies on regulation of its metabolism. Here we performed the first integrated data analysis of *Y. lipolytica* grown in carbon and nitrogen limited chemostat cultures. We first reconstructed a genome-scale metabolic model and used this for integrative analysis of multilevel omics data. Metabolite profiling and lipidomics was used to quantify the cellular physiology, while regulatory changes were measured using RNAseq. Analysis of the data showed that lipid accumulation in *Y. lipolytica* does not involve transcriptional regulation of lipid metabolism but is associated with regulation of amino-acid biosynthesis, resulting in redirection of carbon flux during nitrogen limitation from amino acids to lipids. Lipid accumulation in *Y. lipolytica* at nitrogen limitation is similar to the overflow metabolism observed in many other microorganisms, e.g. ethanol production by *Saccharomyces cerevisiae* at nitrogen limitation.

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INTRODUCTION

The yeast *Yarrowia lipolytica* has a high potential as microbial cell factory for the production of biofuels and chemicals. *Y. lipolytica* is an oleaginous yeast, capable of accumulating over 70% of its biomass as lipids.¹ These lipids are stored in lipid bodies and exist primarily of triacylglycerols (TAGs) with different chain lengths, which can function as intermediates for the production of advanced biofuels.² Studying metabolism in *Y. lipolytica* is of interest as its dysregulation allows engineering opportunity for increased lipid production. As an example, a push-and-pull genetic engineering strategy has been employed to divert the carbon flux during nitrogen restriction towards TAG production.³ Overexpression of acetyl-CoA carboxylase (*ACC1*) pulls carbons from the typical excretion metabolite citrate into fatty acid biosynthesis, while overexpression of diacylglycerol acyltransferase (*DGA1*) pushes the fatty acids into the TAGs. This approach has been successful as almost all carbons are diverted from citrate excretion into TAG biosynthesis. However, while efforts have been made to increase lipid production in *Y. lipolytica*,⁴ currently our knowledge of how lipid accumulation is regulated in *Y. lipolytica* is limited, whereas regulators as *SNF1*,⁵ *MIG1*⁶ and *MGA2*⁷ have been shown to affect lipid accumulation. Systems level analysis is an excellent tool for probing regulatory mechanisms, as demonstrated extensively for *Saccharomyces cerevisiae*.^{8,9} However no such approach has been applied to *Y. lipolytica* to date, and we therefore undertook the first integrated analysis of lipid accumulation in *Y. lipolytica*.

RESULTS

Reconstruction of genome-scale metabolic model

Integration of multilevel data requires a framework that can accommodate different data types and allows for mapping of

interactions. For this purpose we generated a comprehensive genome-scale metabolic model (GEM) of *Y. lipolytica* metabolism. Although three GEMs of *Y. lipolytica* have been published previously (iYL619_PCP,¹⁰ iNL895¹¹ and iMK735¹²), our model is far more comprehensive, based on the more recent Yeast 7.11 consensus network¹³ and curated to include unique reactions from both iYL619_PCP and iNL895. Further curation was performed with available literature data and improved annotation of the *Y. lipolytica* genome as described in Supplementary Informations 1 and 2. The resulting model is the most comprehensive GEM of an oleaginous yeast to date, and provides the biofuel research community with a tool for further identifying engineering targets and a framework to unravel regulation of metabolism. We name this model *iYali4*, as the fourth published GEM of *Y. lipolytica*.

Physiological characterisation of chemostat cultures

A high-lipid producing strain of *Y. lipolytica*, overexpressing diacylglycerol acyltransferase (*DGA1*), the last step of TAG biosynthesis, was cultivated in a bioreactor under chemostat conditions. Restrictive availability of ammonium (as nitrogen source) was compared to the restrictive availability of glucose (as carbon source), all at the same dilution rate (0.05 per h). Carbon–nitrogen ratios of 2.2 and 110 were selected as representing carbon versus nitrogen limitation (Supplementary Information 3). When the cultures reached steady state, defined at a constant OD₆₀₀ and O₂ partial pressure in the exhaust gas, samples were taken for measurements (Figure 1).

An overview of the fermentation profile shows that the specific glucose consumption rate (q_{gluc}) was similar at both nitrogen and carbon limitations, and also the biomass yield (Y_{SX}) was comparable (Table 1). This is in stark contrast with for instance

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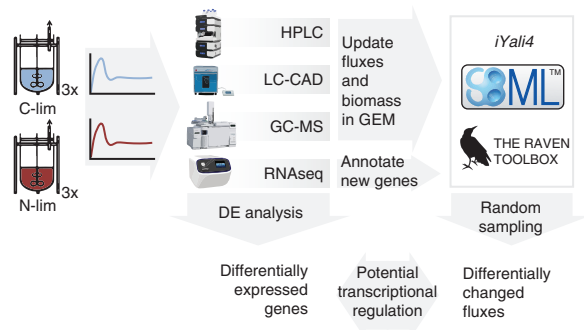


Figure 1. Experimental design. Triplicate chemostats were run at steady-state, after which various samples were taken. Strain-specific GEMs were generated using the experimental data and the predicted flux changes were compared with differentially expressed transcripts, indicating potential transcriptional regulation.

Table 1. Physiological parameters

	Carbon restriction	Nitrogen restriction
Specific growth rate (per hour)	0.047 (± 0.004)	0.048 (± 0.002)
Biomass concentration (g/l)	2.1 (± 0.1)	2.7 (± 0.5)
Extracellular glucose concentration (g/l)	0	17.9 (± 0.8)
q_{Gluc} (mmol gDW/h)	0.61 (± 0.06)	0.64 (± 0.06)
q_{O_2} (mmol gDW/h)	1.3 (± 0.3)	2.1 (± 0.3)
q_{CO_2} (mmol gDW/h)	1.5 (± 0.1)	2.2 (± 0.3)
RQ (–)	1.15 (± 0.20)	1.0 (± 0.1)
Y_{SX} (gDW/g glucose)	0.43 (± 0.02)	0.42 (± 0.01)
Dissolved oxygen (%)	82 (± 4)	74 (± 10)

Abbreviation: RQ, respiratory quotient.

Data are means (s.d.) from three independent chemostats.

S. cerevisiae where nitrogen limitation results in ethanol fermentation and a fivefold decrease in the biomass yield.¹⁴ *Y. lipolytica* is, however, an obligatory aerobe and does not ferment ethanol. Although wild-type *Y. lipolytica* produces citrate as a carbon overflow metabolite, the *DGA1* strain used in this study instead shuttles citrate towards lipid production.³ HPLC analyses of the cell-free supernatants confirmed the absence of citrate excretion. The specific oxygen and carbon dioxide exchange rates are higher during nitrogen restriction (Table 1), suggesting a more active oxidative phosphorylation.

A high degree of filamentous growth (Supplementary Information 4) was observed in all chemostat cultures and shake flask cultivations using the same growth medium, albeit at lower levels in shake flasks. Filamentation has been observed previously in bioreactor cultivations for studies of lipid accumulation.¹⁵ Filamentation can be induced by a range of environmental changes, including the carbon source,^{16,17} nitrogen source,^{16,18,19} temperature,²⁰ oxygenation,^{18,21} pH and buffer composition.^{16,19} In addition, conflicting conclusions can be found in the literature, possible due to strain specificity.²² We adjusted several of these operational parameters in our chemostat fermentations, but none of these adjustments resulted in a major reduction in filamentation, showing that filamentous growth is a complex response that can involve multiple triggers. We did not evaluate the addition of a complete supplement mixture or addition of amino acids to the medium as this would complicate restricting the nitrogen source and complicate quantification of uptake fluxes. Nonetheless, as filamentation occurred at all growth conditions, physiological responses associated with filamentous growth will be filtered out in a comparative analysis.

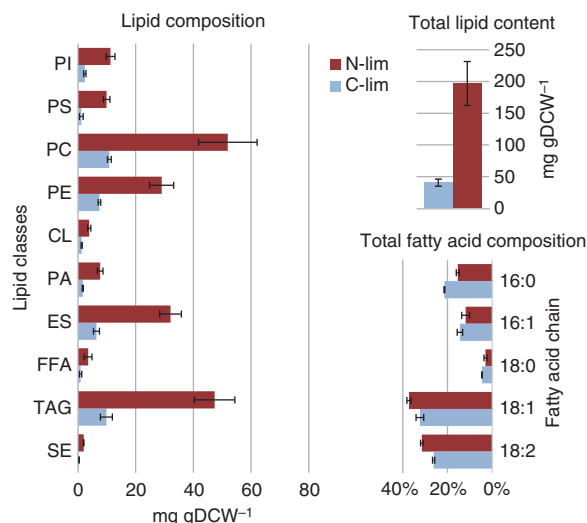


Figure 2. Lipid and total fatty acid composition. Error bars are s.d. of three independent biological replicates. CL, cardiolipin; ES, ergosterol; FFA, free fatty acid; PA, phosphatidate; PC, phosphatidyl-choline; PE, phosphatidylethanolamine; PI, phosphatidyl-inositol; PS, phosphatidyl-serine; SE, steryl ester; TAG, triacylglycerol.

Increases in lipids during nitrogen restriction

The lipid compositions of *Y. lipolytica* were measured for both nutrient restrictions, and the most dominant phospholipids were found to be PE, PC and PI (Figure 2), corroborating previous measurements of *Y. lipolytica*.²³ The low steryl ester (SE) content was surprising, as SE is typically identified as a storage lipid together with TAG, and lipid droplets normally contain 7.8–14% steryl esters.²⁴ However, other reports state that steryl esters are only present in very small amounts.²⁵

The total lipid content increased from 40.5 mg/gDW (± 5.7 mg/gDW) during carbon limitation to 197 mg/gDW (± 35 mg/gDW) during nitrogen limitation (Figure 2). During batch fermentation of the DGA strain the total lipid content reached 338 mg/gDW confirming that the chemostat cultivations simulate conditions occurring earlier in the batch fermentation, when lipids are still actively accumulating.³

The increased lipid content during nitrogen limitation could surprisingly be contributed to an increase in virtually all lipid species and not just triacylglycerols and steryl esters (Figure 2). In *S. cerevisiae* lipid droplets are likely structured with an inner-core of TAGs, surrounded by a shell of steryl esters and an outer surface of a phospholipid monolayer,²⁶ whereas lipid droplets in *Y. lipolytica* grow in size when more lipids are accumulated.²⁴ Larger lipid droplets require more total phospholipids, whereas the ratio of phospholipid per lipid content would decrease, albeit they only contribute 1.6–2% to the total amount of lipids in lipid droplets.²⁴ The increase could also be due to a larger amounts of other intracellular membranes. The increase in filamentation (Supplementary Information 4) observed during nitrogen limitation may contribute to the increased phospholipids. A more cylindrical shape of the cells result in a larger surface-to-volume ratio, whereas the amount of lipids in the cell wall remains at 5% in both yeast and filamentous *Y. lipolytica*.²⁷

All isolated lipids were derivatized to release fatty acid methyl esters (FAMES) that were subsequently analysed by GC-MS. The fatty acid composition shows only a minor shift towards shorter chain lengths (Figure 2); C16 versus C18; and the saturation level decreases at nitrogen limitation compared with carbon limitation. This is in contrast to batch fermentations where C18:0 is the second most prominent fatty acid,³ whereas C16:1 and C18:2 have the lowest levels.³

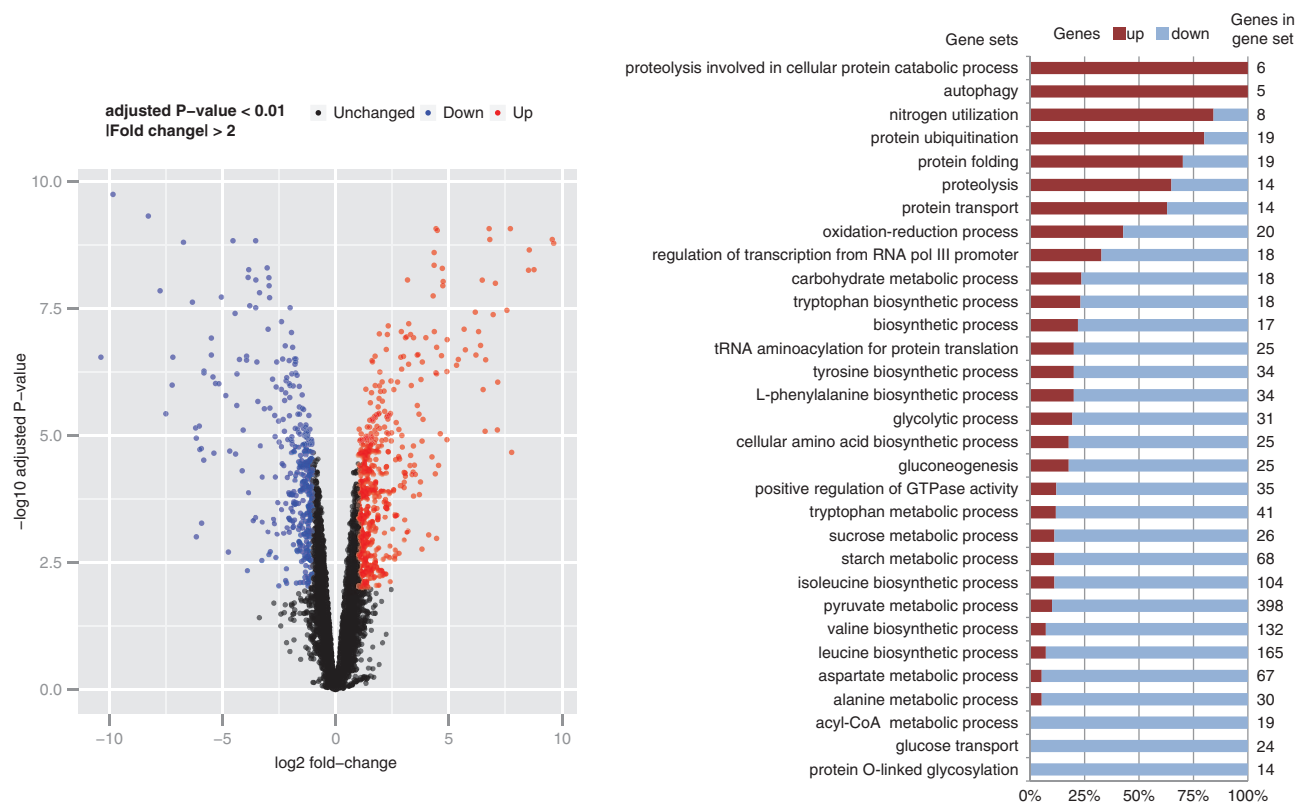


Figure 3. Gene-set analysis of nitrogen over carbon limitation. Carbon limitation functioned as reference condition. Gene sets are defined by GO terms, and for significantly enriched GO terms (adjusted $P < 0.01$) is shown what percentage of genes are either up- or downregulated at nitrogen limitation. The top GO terms are upregulated while the lower GO terms are downregulated.

Transcriptional changes during nitrogen limitation

To quantify the regulatory responses to the shift in nutrient limitation we performed RNA sequencing. Gene-set analysis was performed on normalised RNA counts to obtain a systemic overview of the differential expression, where gene-sets were defined by GO term annotations obtained from various databases. The overall picture obtained was that *Y. lipolytica* under nitrogen limitation attempts to minimise its nitrogen usage (Figure 3). GO terms enriched for transcripts that are upregulated during nitrogen limitation are involved in protein turnover and autophagy, providing alternative sources of nitrogen. In contrast, many of GO terms enriched for downregulated transcripts are related to amino-acid metabolism.

The transcriptional response appears highly regulated with about 30 putative transcription factors differentially expressed (Supplementary Information 5). Although transcriptional regulatory networks can be subjected to rewiring,²⁸ such as the shift of SREBP from regulating ERG genes to regulate filamentation,²⁹ other parts of transcriptional regulatory networks are more conserved.

Several signalling networks are involved in the response to use of different nitrogen sources in *S. cerevisiae*.³⁰ Nitrogen restriction releases nitrogen catabolite repression (NCR), which is regulated by interplay of four GATA transcription factors conserved to *Y. lipolytica*, Gln3, Gat1, Gzf3 and Dal80. Homology prediction is difficult for their genes due to high similarity, however, Yali0D20482g and Yali0C22682g are likely GAT1 and GZF3, whereas GLN3 and DAL80 are Yali0F17886g and Yali0E05555g.³¹ GAT1 and GZF3 are strongly upregulated during nitrogen limitation (Supplementary Information 5). Interestingly, glutamate dehydrogenase (Yali0F17820g; Yali0E09603g), glutamate synthase (Yali0B19998g) and glutamine synthetase (Yali0F00506g) are not upregulated during nitrogen limitation, while they are

upregulated in nitrogen restricted *R. toruloides*.³² As an alternative route to recycle nitrogen, *Y. lipolytica* has an ortholog (Yali0A06974g) of the *Aspergillus nidulans* xanthine dehydrogenase,³³ which is absent in *S. cerevisiae*. Presence of this pathway means that purines can be degraded via allantoin to release ammonia. Multiple genes in this pathway are under control by Gat1 and Gln3, whereas this whole pathway is upregulated during nitrogen restriction (Figure 3), corroborating the release of NCR.

A more central role in nitrogen sensing and signalling is played by the TOR complex,³⁴ where nitrogen starvation inhibits TOR, what subsequently, among others, inhibits protein translation, activates autophagy and reconfigures amino-acid permeases.³⁰ The observed differential gene expression profile supports TOR-mediated regulation, such as upregulation of autophagy, proteasome, peptidases and ubiquitination (Figure 3).

Another key regulator of metabolism is Snf1, a protein kinase that is involved in many signalling pathways, including the shift from fermentation to aerobic metabolism in *S. cerevisiae*, regulation of amino-acid metabolism, inhibition of translation and activation of β -oxidation. A *Y. lipolytica* knockout of SNF1, or other components of the Snf1 signalling complex, has an increased lipid content, and from this it has been proposed that Snf1 is an important regulator of lipid accumulation in *Y. lipolytica*.⁵ In *S. cerevisiae* Snf1 inhibits the activity of the acetyl-CoA carboxylase Acc1 by phosphorylation,³⁵ and these phosphorylation-sites on Acc1 are conserved in *Y. lipolytica* Acc1. However, the gene expression profile of the SNF1 knockout was very distinct from the chemostat expression profiles, even though both presented an oleaginous phenotype.⁵ For instance, removal of Snf1 increases expression of β -oxidation⁵ while this pathway was downregulated during nitrogen restricted chemostat culture, represented by the GO terms acyl-CoA dehydrogenase and

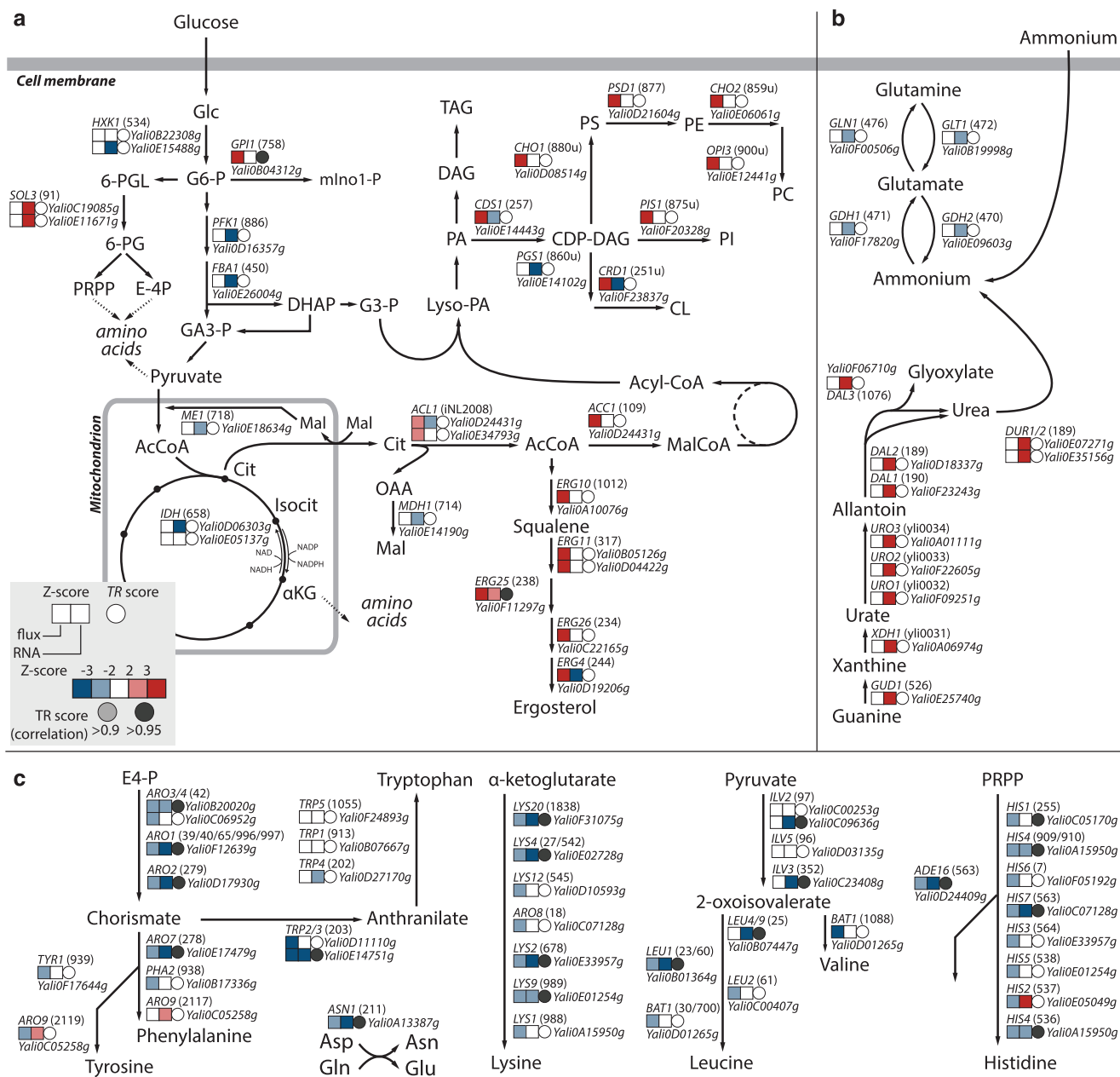


Figure 4. Schematic overview of changes in flux and transcript levels. Arrows can represent multiple reactions. Z-scores indicate changes on the levels of metabolic flux or RNA, TR score indicates a correlation between metabolic flux and RNA, suggesting transcriptional regulation. (a) Central carbon metabolism including lipid metabolism. (b) Nitrogen metabolism. (c) Amino acid biosynthetic pathways.

oxidase (Figure 3). β -oxidation could be considered as a futile cycle during lipid accumulation, although it has been reported that *Y. lipolytica* degrades its reserves lipids even in presence of high carbon concentration in the medium.^{36,37} *Y. lipolytica* has six acyl-CoA oxidases with different specificities.³⁸ The deletion of all six genes produces a higher lipid content²⁵ while deletion of the peroxisomal biogenesis gene *PEX10* also increases lipid accumulation.⁴ These metabolic engineering strategies are based on manipulating the β -oxidation response of *Y. lipolytica* in oleaginous conditions. This questions the important role of Snf1 in the regulation of lipid metabolism. The *SNF1* knockout is forced to accumulate lipids due to the lack of repression of *Acc1* by Snf1, however, the observed transcriptional response is not

representative for lipid accumulation, and therefore the role of Snf1 in lipid accumulation is likely limited to its regulation of *Acc1*.

Correlation of transcription and metabolic fluxes

Although differential expression of genes is indicative of how the organism reacts to a certain treatment or condition, changes in transcript levels do not necessarily translate to changes in protein levels and, when these proteins are enzymes, in changes in metabolic fluxes. Nonetheless, correlations of transcriptional changes with changes in metabolic fluxes suggest transcriptional regulation of those reactions. Therefore, metabolic fluxes for each reaction were estimated using the *Y. lipolytica* GEM, by constraining the model with condition-specific experimental data, i.e., glucose and oxygen consumption rates, CO_2 production rate, growth rate and measured lipid composition (Table 1, Figure 2).

The resulting models were sampled using a random sampling algorithm,³⁹ providing means and s.d. for each flux in either nitrogen or carbon limitation. Standard scores were calculated for each reaction and these were compared with standard scores calculated for the transcriptional changes. Of the 2,194 gene-reaction pairs, 53 showed transcriptional regulation (Supplementary Information 6).

Our gene-set analysis already identified downregulation of amino-acid metabolism (Figure 3). This downregulation is also observed in the metabolic fluxes through several of the amino-acid biosynthetic pathways (Figure 4). Specifically, leucine has been suggested to be involved in the regulation of lipid metabolism in *Y. lipolytica*.⁴ The *LEU2* gene is an often used selective marker on overexpression plasmids to metabolically engineer leucine auxotrophic strains (e.g., Po1g), while the additional supplementation with leucine, and not isoleucine, resulted in a higher accumulation of lipids.⁴ It is plausible that *Y. lipolytica* senses the increased leucine level and as a transcriptional response further downregulates amino-acid biosynthesis. This further downregulation then provides an ever stronger redirection of flux towards lipid metabolism. Interestingly, in *S. cerevisiae* leucyl-tRNA synthetase has been identified as controlling TORC1 in a leucine-dependent manner.⁴⁰ Although this identifies an intracellular leucine sensor, the activation of TORC1 with increased leucine is actually the opposite as what is observed during nitrogen limitation.

Lipid metabolism has limited transcriptional regulation

While the lipid contents of the cells change drastically upon nitrogen limitation, the transcript levels of lipid metabolism was poorly regulated. In contrast to the increasing lipid content at nitrogen limitation, some of the transcripts were downregulated at this condition. Lipid metabolism therefore does not appear to be regulated at the transcriptional level.

Previously, the transcriptional response to the onset of lipid accumulation has been studied in fed-batch cultivations.³¹ There are numerous differences observed in expressional changes between the two data sets, including changes in isocitrate lyase, aconitase and dephospho-CoA kinase expression during fed-batch cultivations that are not changing in the chemostats. Many of these difference can be explained by the fact that the chemostat experiments only compares steady-state nitrogen versus carbon limitation, whereas the fed-batch cultivation includes adaptation and growth-related differences. Nonetheless, both data sets show similar trends, such as changes related to NCR, but more importantly the lack of transcriptional regulation of lipid metabolism, not only during steady-state lipid accumulation but also during its transition towards this state.³¹

While metabolic engineering efforts have been made in the overexpression of genes part of the lipid biosynthetic pathway,^{3,15} it seems that these strategies are based on crudely forcing the flux towards lipid metabolism instead of taking the native regulatory mechanisms into account. In contrast, the native regulation that occurs during nitrogen limitation is primarily focused on regulating amino-acid metabolism (Figure 5).

DISCUSSION

Limitation of nitrogen during continued growth of *Y. lipolytica* provokes a dramatic response in the biomass composition, i.e. a sustained increase in virtually all lipids. It has been proposed that an important role in this process is the inhibition of AMP-dependent isocitrate dehydrogenase evoked by an increased activity of AMP deaminase.⁴¹ Although transcriptional regulation likely has another important role in this response, as not only a significant number of transcripts but also many TFs are differently expressed, it appears that lipid metabolism itself is undergoing

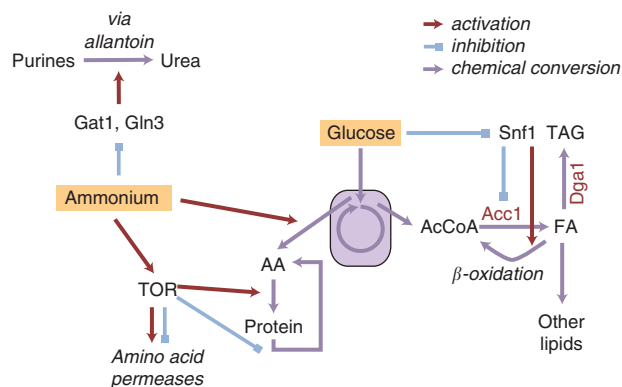


Figure 5. Schematic overview of regulation that occurs during nitrogen limitation in a high-lipid producing strain.

very limited transcriptional regulation. The metabolic network is adapted to divert the carbon flux from pathways requiring nitrogen, such as amino-acid metabolism, while the lipid biosynthetic pathways are seemingly in high enough activity to harbour this increased flux. Nonetheless, it is likely that additional regulation takes place such as post-translational modifications, as it is known that Acc1 activity is repressed by phosphorylation by Snf1, which probably contributes to the oleaginous phenotype in the *SNF1*-knockout. This corroborates findings from fed-batch cultivations of *Y. lipolytica*, where also limited transcriptional regulation of lipid metabolism was observed.³¹ In addition, in *R. toruloides* the limited transcriptional regulation of lipid metabolism is primarily focused on the downregulation of β -oxidation.³² Although *Y. lipolytica* does not seem to regulate the expression of its lipid metabolism, metabolic engineering efforts have been made to increase the lipid yield by overexpression of additional copies of native genes involved in lipid metabolism, such as *ACC1*, *DGA1* and recently stearoyl-CoA desaturase.¹⁵ These efforts have been successful; however, it ignores the original regulatory mechanisms of the cell. Taking the native regulatory network into account will result in more robust and increased lipid yields, and our findings may enable coupling growth and lipid accumulation, which is required for obtaining high specific productivities of TAG.

A more systematic study of lipid accumulation in oleaginous yeasts requires a high-quality framework to integrate the various data, which we provide here with a high-quality genome-scale model of *Y. lipolytica*. Our findings point to that Snf1 might not naturally have an important role in lipid accumulation as previously postulated. Thus, even though a knockout of *SNF1* results in an oleaginous phenotype, the transcription profile is the opposite of what happens during lipid accumulation induced by nitrogen restriction. Our results therefore point to that lipid accumulation in *Y. lipolytica* at nitrogen limitation is similar to the overflow metabolism observed in many other microorganisms, e.g. ethanol production by *S. cerevisiae* at nitrogen limitation, and more in line with what is observed in mammals where a high carbon to nitrogen ratio in the diet results in TAG accumulation.

MATERIALS AND METHODS

Yeast strain and chemostats

The *Yarrowia lipolytica* strain used in this study was derived from Po1g (Leu⁻), contains the MTYL053 plasmid expressing *DGA1* and was provided by ref. 3. Chemostat cultures were performed with a dilution rate of 0.05 per hour at 30 °C in 1.2 l bioreactors (DASGIP, Jülich, Germany) with a working volume of 750 ml at pH 3.5, controlled with 2 M KOH. Dissolved oxygen was kept above 30% with a stirrer rate of 600 rpm and an air flow rate of 1 v.v.m. For carbon restriction, 1 l medium contained 5 g glucose, 5 g (NH₄)₂SO₄, 3 g KH₂PO₄, 0.5 g MgSO₄·7 H₂O, vitamins and trace metal

solutions⁴² and 125 µl antifoam 204 (Sigma-Aldrich, St Louis, MO, USA). For nitrogen restriction a similar medium was used, where the glucose was increased to 25 g, the (NH₄)₂SO₄ decreased to 0.5 g while SO₄ levels remained constant by adding 5.96 g K₂SO₄. CO₂ and residual O₂ in the exhaust gas were measured using an online gas analyser (DASGIP). Samples for cell dry weight, lipid and FAME analysis and transcriptomics were taken when the chemostats reached steady state, defined as stable CO₂ and O₂ outflow and optical density, which was achieved after circa 120 h.

Lipid and fatty acid analysis

Samples were taken from the steady-state chemostat cultivations to investigate their lipid contents and compositions. The biomass was disrupted and lipids extracted using a fast microwave-assisted extraction method,⁴³ and subsequently analysed by LC-CAD, allowing quantification of 10 different lipid species. Fatty acids were extracted and derivatized to fatty acid methyl esters using a similar microwave-assisted method⁴⁴ and subsequently analysed by GC-MS.

RNA extraction and sequencing

Samples for RNA analysis were rapidly taken from steady-state chemostats, frozen in liquid nitrogen and stored at −80 °C. Total RNA was extracted with Trizol (Invitrogen, Carlsbad, CA, USA) using the manufacturer's instructions with additional mechanical disruption of the cells using a FastPrep homogenizer (MP Biomedicals, Santa Ana, CA, USA) and 1 mm silica beads. Further RNA preparation and RNA sequencing was performed by SciLifeLab in Uppsala, Sweden using their IonTorrent platform. Data are deposited at ArrayExpress (E-MTAB-3837). Raw RNAseq reads were aligned to the *Y. lipolytica* CLIB122 reference genome using local alignment in Bowtie,⁴⁵ counts were obtained with HTSeq⁴⁶ and transformed using voom.⁴⁷ In all analyses, carbon limitation was set as reference condition.

De novo assembly

Raw RNA data from all samples was concatenated and reads of < 40 base pairs were removed. Transcripts were *de novo* assembled and coding sequences predicted using the Trinity software package.⁴⁸ Proteins were functionally annotated using Blast2GO.⁴⁹ Full length transcripts and coding sequences were aligned to the reference genome contigs from strain CLIB122⁵⁰ using Blat⁵¹ to determine their position and percent identity. Determination of best hits was made using Identity score, calculated as [(Q end—Q start—Q gap bases)/Q size+(T end—T start—T gap bases)/T size] × [match/(match+mis-match)]/2. Transcripts with an identity score < 0.4 were considered poor quality alignments.

Genome-scale modelling

A genome-scale model of *Y. lipolytica* was reconstructed using the RAVEN toolbox in Matlab⁵² and the yeast consensus network version 7.11 (ref. 13) was employed as a template. To facilitate integration of our FAME analysis, we reverted the extensive expansion of lipid metabolism in yeast 7, where the length of each acyl-chain is specified. A previously published *Yarrowia* GEM was assessed and curation on this model were transferred to our model.¹¹ Additional gap filling and curation was performed manually. This resulted in our model *iYali4*, available from BioModels database (MODEL1508190002) and as Supplementary Information 7.

Flux ranges were determined using the random sampling approach,³⁹ where random combinations of objective functions are used to sample the edges of the solution space. The difference in flux ranges between carbon and nitrogen restricted conditions were converted to Z-scores, and likewise for the mRNA fold changes, as described previously.³⁹

Gene set analysis

Gene sets were generated from GO term annotation of the *Yarrowia* genome as provided by Blast2GO,⁴⁹ Ensembl,⁵³ InterPro⁵⁴ and UniProt.⁵⁵ A total of 29,513 GO terms were annotated to 5,232 genes. Gene set analysis was performed with the normalised RNAseq counts using Piano,⁵⁶ with the mean of the gene-level statistics, ignoring gene-sets smaller than 5 and larger than 500 genes, sampling 1,000 times and corrected for multiple testing using FDR.

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COMPETING INTERESTS

The authors declare no conflict of interest.

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